

The complete mitochondrial genome of the parthenogenetic Caucasian rock lizard *Darevskia unisexualis* (*Squamata: lacertidae*) contains long tandem repeat formed by 59 bp monomer

Aleksey Komissarov^a, Vitaly Korchagin^b, Sergei Kliver^a, Pavel Dobrynin^a, Serafima Semyenova^b, Andrey Vergun^b , Stephen O'Brien^a and Alexey Ryskov^b

^aTheodosius Dobzhansky Center for Genome Bioinformatics Saint Petersburg State University, St. Petersburg, Russia; ^bLaboratory of Genome Organization, Institute of Gene Biology of the Russian Academy of Sciences, Moscow, Russia

ABSTRACT

The first complete mitochondrial genome sequence of parthenogenetic Caucasian rock lizard *Darevskia unisexualis* (*Lacertidae* family) is determined by hybrid assembly with Illumina HiSeq and PacBio RS II platforms. The circular 21.4 kbp mitogenome contains 13 protein-coding genes, 12S and 16S rRNA genes, 20 tRNAs, two pseudogenized tRNAs, and one long tandem repeats with 4.1 kbp length formed by 59 bp monomer repeated x70.6 times located before control region. This finding represents a new example of mitogenome variation in lizards of hybrid origin, providing fundamental data for following study of a unique hybridization system formed by parthenogenetic and bisexual species in the mountain steppe of central Armenia.

ARTICLE HISTORY

Received 22 September 2016
Accepted 23 October 2016

KEYWORDS

Mitochondrial genome; parthenogenetic lizards; rock lizards; *Darevskia unisexualis*; tandem repeats

Parthenogenetic lizard species represent a unique model of organisms for study of genetic and ecological bases of hybridogeneous speciation. *Darevskia unisexualis* is one of the seven parthenogenetic species originated from the interspecific hybridization of *D. raddei nairensis* (maternal species) and *D. valentini* (paternal species), which belong to different clades (Moritz et al. 1992; Murphy et al. 2000; Fujita & Moritz 2010).

Mitochondrial genomes of *Squamata* often have unusual and complicated structure (Amer and Kumazawa 2005; Dong & Kumazawa 2005; Amer & Kumazawa 2007; Fujita et al. 2007; Kumazawa et al. 2014). In this article we report a new kind of rearrangement in mitochondrial genome of asexual lizard *Darevskia unisexualis*: a long tandem repeat with 59 bp monomer located before the control region (Darevsky et al. 1985; Moritz et al. 1992; Murphy et al. 2000; Fujita & Moritz 2010).

DNA samples extracted from blood of *D. unisexualis* obtained previously between 1997 and 2006 from lizards of Armenian population Takyarlu (Artavaz, 40°37'20.2"N 44°34'51.4"E) were delivered to the Aseq/Macrogen (Korea) for genomic library preparation and sequencing with Illumina HiSeq 2000 and PacBio RS II.

To overcome complexity of control region assembly we used approach based on reads extraction with following greedy assembly previously successfully adopted in *Solenodon paradoxus* mitogenome assembly (Brandt et al. 2016). A subset of Illumina reads having common kmers with *Podarcis siculus* mitogenome (NC_011609; Podnar et al. 2009)

was extracted with Cookiecutter software (Starostina et al. 2015), mapped back to the reference mitogenome with bowtie2 (Langmead & Salzberg 2012), and assembled into consensus sequence, that was used as query for alignment against PacBio reads with Blasr aligner (Chaisson & Tesler 2012). Cookiecutter's kmer database was updated from 215 successfully found PacBio reads. Updated kmer database was used for accurate read extraction from Illumina HiSeq raw data. After that Illumina reads were mapped back to PacBio reads and assembled to the final sequence. The consistency of assembly was verified: (1) by consistency with PacBio reads that were mapped to it with MAFFT tool (Katoh & Standley 2013); (2) by consistency with previously published restriction map (Moritz et al. 1992) that has a perfect match with assembled mitogenome except two inverted restriction sites; (3) consistency of Illumina reads coverage. The assembled genome was annotated with MITOS WebServer (Bernt et al. 2013) with additional checking for predicted tRNA and protein-coding genes.

The circular assembled mitogenome size is 21,433 bp, with 13 predicted protein-coding genes (PCGs), two ribosomal RNA (rRNA) genes, and 20 tRNAs, two pseudogenized tRNAs, and one complex tandem repeats with 59 bp monomer repeated x70.6 times and located before control region. MITOS WebServer predict second ND4 in 17202–18491 regions inside found tandem repeat, however, verification by Blast alignment did not find any significant match in this region. The validation of phylogenetic position is shown in Figure 1.

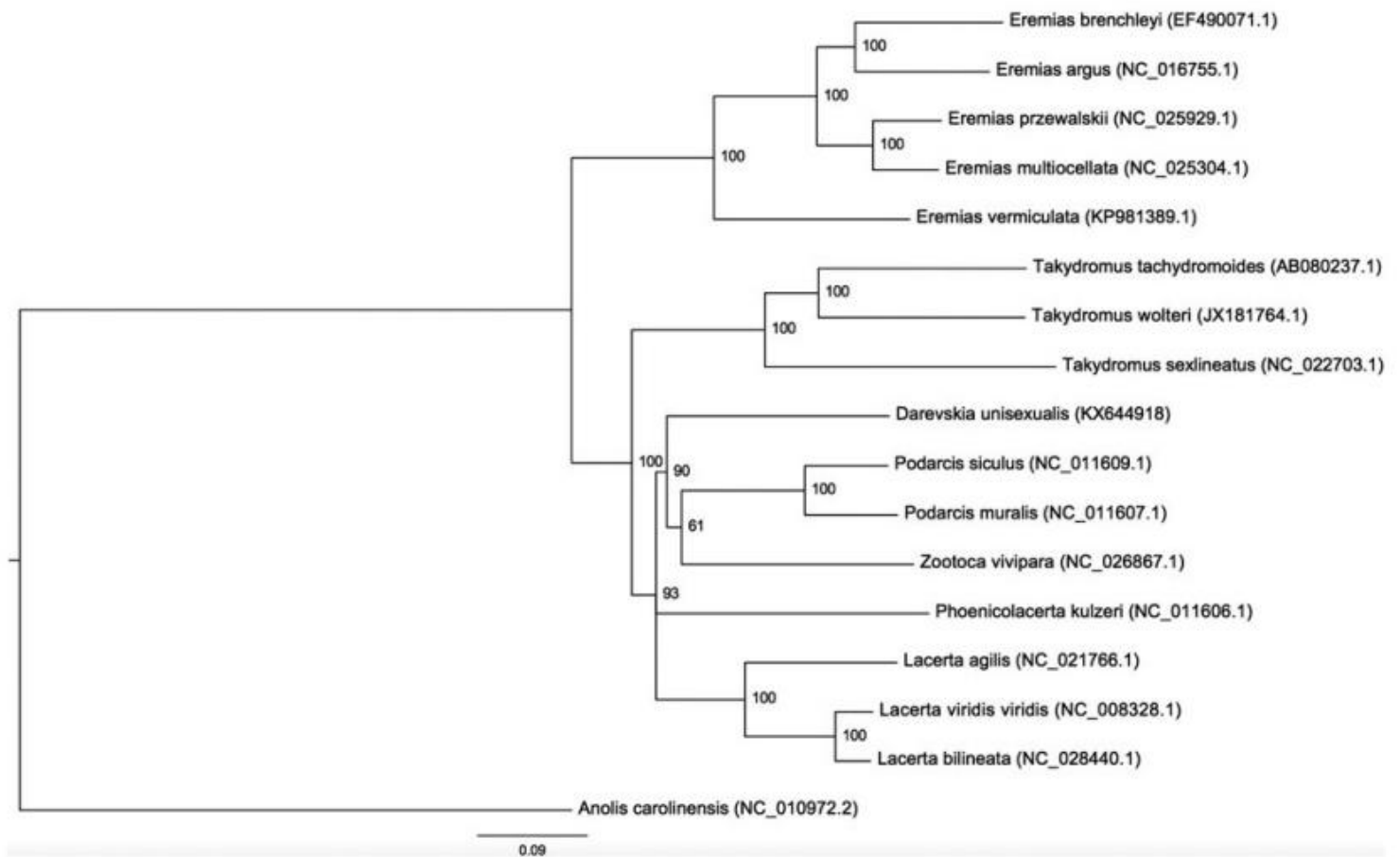


Figure 1. Molecular phylogeny of *Daresvskia unisexualis* and other Lacertidae species based on all mitochondrial protein-coding genes and rRNAs. Each of alignments was performed separately by MAFFT v7.187 (Katoh & Standley 2013), then alignments were concatenated. Tree was reconstructed by RAxML v8.0.22 (Stamatakis 2014) with 10,000 bootstrap replicates. All nodes with support lower than 50 were removed using ETE toolkit (Huerta-Cepas et al. 2010) and final tree was drawn by FigTree (<http://tree.bio.ed.ac.uk/software/figtree/>).

Unusual mitochondrial genome of parthenogenetic lizards containing long tandem repeat raises questions about stability and evolution of mtDNA after recent hybridization events. Moreover, its features provide unique opportunities to study mtDNA evolution not only in *Squamata* taxon, but in all animals.

Data availability

D. unisexualis mitogenome assembly is available from Genbank with accession number KX644918.

Disclosure statement

The authors declare that they have no conflicts of interest concerning this article.

Funding

This study was supported by the Russian Science Foundation, 10.13039/501100006769 [14-14-00832]

ORCID

Andrey Vergun  <http://orcid.org/0000-0001-5255-7802>

References

Amer SA, Kumazawa Y. 2005. Mitochondrial genome of *Pogona vitticeps* (Reptilia; Agamidae): control region duplication and the origin of Australasian agamids. *Gene*. 346:249–256.

- Amer SA, Kumazawa Y. 2007. The mitochondrial genome of the lizard *Calotes versicolor* and a novel gene inversion in South Asian dracoline agamids. *Mol Biol Evol*. 24:1330–1339.
- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritsch G, Pütz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo metazoan mitochondrial genome annotation. *Mol Phylogenet Evol*. 69:313–319.
- Brandt AL, Grigorev K, Afanador-Hernández YM, Paulino LA, Murphy WJ, Núñez A, Komissarov A, Brandt JR, Dobrynin P, Hernández-Martich JD, María R. 2016. Mitogenomic sequences support a north–south subspecies subdivision within *Solenodon paradoxus*. *Mitochondrial DNA Part A*. 14:1–9.
- Chaisson MJ, Tesler G. 2012. Mapping single molecule sequencing reads using basic local alignment with successive refinement (BLASR): application and theory. *BMC Bioinform*. 13:238.
- Darevsky IS, Kupriyanova LA, Uzzell T. 1985. Parthenogenesis in reptiles. In: Gans C, Billett DF, editors. *Biology of Reptilia*. vol. 15. New York: John Wiley & Sons; p. 413–526.
- Dong S, Kumazawa Y. 2005. Complete mitochondrial DNA sequences of six snakes: phylogenetic relationships and molecular evolution of genomic features. *J Mol Evol*. 61:12–22.
- Fujita MK, Boore JL, Moritz C. 2007. Multiple origins and rapid evolution of duplicated mitochondrial genes in parthenogenetic geckos (*Heteronotia binoei*; *Squamata*, *Gekkonidae*). *Mol Biol Evol*. 24:2775–2786.
- Fujita MK, Moritz C. 2010. Origin and evolution of parthenogenetic genomes in lizards: current state and future directions. *Cytogenet Genome Res*. 127:261–272.
- Huerta-Cepas J, Dopazo J, Gabaldón T. 2010. ETE: a python environment for tree exploration. *BMC Bioinform*. 11:1.
- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol*. 30:772–780.
- Kumazawa Y, Miura S, Yamada C, Hashiguchi Y. 2014. Gene rearrangements in gekkonid mitochondrial genomes with shuffling, loss, and reassignment of tRNA genes. *BMC Genom*. 15:1.

- Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. *Nat Methods*. 9:357–359.
- Moritz C, Uzzell T, Spolsky C, Hotz H, Darevsky I, Kupriyanova L, Danielyan F. 1992. The material ancestry and approximate age of parthenogenetic species of Caucasian rock lizards (Lacerta: Lacertidae). *Genetica*. 87:53–62.
- Murphy RW, Fu J, Macculloch RD, Darevsky IS, Kupriyanova LA. 2000. A fine line between sex and unisexuality: the phylogenetic constraints on parthenogenesis in lacertid lizards. *Zool J Linn Soc*. 130:527–549.
- Podnar M, Pinsker W, Mayer W. 2009. Complete mitochondrial genomes of three lizard species and the systematic position of the Lacertidae (Squamata). *J Zool Syst Evol Res*. 47:35–41.
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics*. 30:1312–1313.
- Starostina E, Tamazian G, Dobrynin P, O'Brien SJ, Komissarov A. 2015. Cookiecutter: a tool for kmer-based read filtering and extraction. *bioRxiv*. Available from: <http://dx.doi.org/10.1101/024679>.